Preparation and Characteristics of Novel Porous PLGA Microsphere by Gas Foaming Method Using Hydrogen Peroxide

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Statement of Purpose: The goal of this study is to fabricate porous biodegradable microcarriers with sustained release for drug delivery. Hydrogen peroxide (H₂O₂), a strong oxidizing agent, is a colorless liquid that is decomposed into water and oxygen when heated over 80°C. It also decomposes in the presence of numerous catalysts, e.g., most metals, acids, or oxidizable organic materials, and catalase. In this study, porous poly(lactideco-glycolide) (PLGA) microspheres were fabricated by a double emulsion solvent evaporation method using H₂O₂ as a gas foaming agent. H₂O₂ was mixed in the polymer emulsion droplets to generate oxygen gas bubbles during the solvent evaporation process. Catalase was used as a H₂O₂ decomposition catalyst. This method is simple and environment-friendly, eliminating the use of salts. Sterilizing effect is also expected from this process. In this study, various conditions were tested and the resultant porous microspheres were characterized.

Methods: Porous PLGA microspheres were fabricated by a water-in-oil-in-water $(W_1/O/W_2)$ double emulsion solvent evaporation method. PLGA was dissolved in methylene chloride as an oil phase. H₂O₂ aqueous solution (W₁ phase) containing different amounts of H₂O₂ was added to a PLGA solution. The first water-in-oil (W₁/O) emulsion was prepared by a homogenizer. This primary emulsion was immediately poured into a beaker containing polyvinyl alcohol (PVA) solution and catalase as a H_2O_2 decomposition catalyst (W_2 phase). The resultant emulsion solution was placed in a hood under a magnetic stirring condition, allowing the solvent to evaporate. The hardened microspheres were centrifuged, washed three times with deionized water and lyophilized using a freeze dryer. The surface and internal structure of porous microsphere were analyzed by scanning electron microscopy (SEM). The size distribution and the average diameter of microspheres were measured by a particle size analyzer.

Results: Porous PLGA microspheres were fabricated by using a $W_1/O/W_2$ double emulsion solvent evaporation method with H_2O_2 aqueous solution as a gas foaming agent (Fig. 1. (c)). The pores were uniformly distributed and well interconnected throughout the inner part of the microsphere. It was demonstrated that H_2O_2 in the W_1 droplets generated oxygen bubbles upon contacting the catalase in the W_2 phase, thereby creating porous morphology throughout the PLGA microspheres. In the case of pure water as the W_1 phase, the surface of the

microsphere had no pores and the inner core had very tiny pores (Figure 1 (b)). One of the interesting points in this study is that when using H_2O_2 as a gas foaming agent to generate a porous PLGA microsphere, the outer surface of the microsphere becomes covered with a thin PLGA membrane. It is postulated that such phenomena happens by chemical reactivity of the H_2O_2 , but the exact causes are still under research. These covered microspheres represent a novel structure with the benefit that when a drug-loaded microsphere is introduced to the human body, it arrives at the target without initial burst release and dissolves the membrane covering the microsphere to create a sustained drug release. Ultimately, covered microspheres have characteristics that can control the release of drugs with more precise and effective behavior. As a result, hydrogen peroxide, as a new environmentfriendly gas foaming agent for porous microspheres, can produce a unique encapsulated structure more rapidly and easily with the benefit of a sterilizing effect.



Figure 1. SEM images of the surface (top) and crosssection (bottom) of microspheres prepared at various condition (x 500): (a) control PLGA, (b) water, (c) H_2O_2 aqueous solution.

Conclusions: Porous PLGA microspheres were successfully prepared by double emulsion solvent evaporation method using H_2O_2 as a gas foaming agent. Therefore, the porous biodegradable PLGA microsphere is applicable to drug and cell delivery as a biodegradable microcarrier with controlled surface encapsulation for sustained drug release.

References:

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