

Reference Scaffolds for Tissue Engineering

Carl G. Simon, Jr., Jirun Sun, Marcus T. Cicerone

Polymers Division, National Institute of Standards and Technology, Gaithersburg, MD, USA

Statement of Purpose: A reference scaffold is being developed for use as a standard during characterization of new scaffold constructs. In this work, the structure and permeability of the reference scaffolds is presented. Freeform fabrication (FFF) was chosen to make the reference scaffolds since this technique affords the most precise control of scaffold structure. Scaffold structure was assessed using X-ray microcomputed tomography (μ CT) and permeability was measured using an automated liquid permeameter. The scaffold structural parameters of porosity and pore size as well as scaffold permeability were found to be very reproducible. The reference scaffolds will be available from National Institute of Standards and Technology in the future to serve as calibration standards during characterization of tissue engineering scaffolding materials.

Methods: Reference scaffolds were fabricated from poly(ϵ -caprolactone) (weight averaged molecular weight 65,000) (PCL) using a precision melt extrusion deposition system [1]. Temperature in the feed chamber was 95 °C while the nozzle was 90 °C. Nozzle diameter was 0.178 mm, nozzle speed was 10 mm/s and flow rate was 0.017 mL/min. Strut diameter was approximately 200 μ m. Scaffolds were fabricated into cylinders with 5 mm height and 20 mm diameter (Fig. 1). Scaffolds with three different strut spacings were fabricated (strut-edge to strut-edge distance; gap width): 200 μ m, 300 μ m and 450 μ m. Scaffold morphology was characterized by μ CT (μ CT 40, Scanco Medical). The microfocus X-ray source was set at 45 kVp and 177 μ A to give a spot size of 5 μ m. The samples were scanned at a 15 μ m voxel resolution with an integration time of 0.2 s. 3D reconstructions were made using Sigma 1.2, Support 2 and Threshold 30. The porosity, pore size and wall thickness were determined by direct distance transformation methods [2]. Scaffold permeability was measured with an automated liquid permeameter (LP-101-A, Porous Materials, Inc.) using oil as the liquid (ASTM Oil Viscosity Reference Standard S6). The rate of fluid flow through the scaffolds was measured over a range of pressures (500 Pa to 2000 Pa) and used to calculate permeability using Darcy's Law.

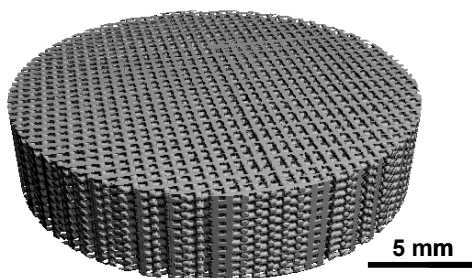


Fig. 1. μ CT image of reference scaffold (300 μ m strut spacing).

Results: Scaffold porosity and pore size increased with increasing strut spacing (Table 1) and differences between

the scaffolds with different strut spacings were significant [$p < 0.05$; ANOVA (analysis of variance) with Tukey's comparison test]. These results fit the expectation that increasing strut spacing will lead to increased porosity and pore size. In addition, the standard deviations for the μ CT-calculated porosity and pore size were $\leq 5\%$, indicating that scaffold fabrication was precise.

Table 1. Scaffold Structural Parameters Determined by μ CT*

Strut Spacing (μ m)	200	300	450
Porosity (%)	44.1 (2.4)	58.5 (3.3)	67.2 (2.0)
Pore Size (μ m)	125 (5)	234 (28)	362 (16)

*Averages ($n = 3$) with standard deviations are given (same as the combined standard uncertainty for the purposes of this work).

The permeability of the scaffolds increased with increasing strut spacing (Fig. 2) and all of the permeabilities for the different strut spacings were significantly different from one another ($P < 0.005$; ANOVA with Tukey's comparison test). These results fit the expectation that increased strut spacing will lead to increased permeability.

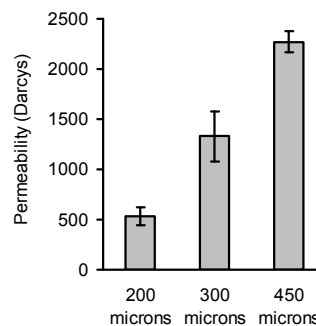


Fig. 2. Permeability of reference scaffolds ($n = 3$; error bars are standard deviation which is the same as the combined standard uncertainty for the purposes of this work).

Conclusions: Freeform fabrication yielded scaffolds with reproducible structural parameters (porosity, pore size) and permeability indicating that the scaffolds could be applied as reference materials for use during characterization of new scaffold materials.

Acknowledgements: We thank Y. Liu, F.W. Wang, J.A. Tesk, F.R. Phelan (all from NIST), D.J. Adams (U. Conn.), L. Shor (Drexel) and Wei Sun (Drexel) for critical input. This work was supported by NIST, NIH/NIBIB R21 EB006497-01 and RESBIO NIH P41 EB001046. This article, a contribution of the National Institute of Standards and Technology, is not subject to US copyright. Certain equipment and instruments or materials are identified in the paper to adequately specify the experimental details. Such identification does not imply recommendation by NIST, nor does it imply the materials are necessarily the best available for the purpose.

References:

- Shor L, Gucer S, Wen X, Gandhi M, Sun W (2007) Biomaterials 28 5291–5297.
- Hildebrand T, Laib A, Muller R, Dequeker J, Ruegsegger P (1999) J Bone Miner Res 14, 1167-1174.