

Novel Nanostructured Coatings Induce Osteogenic Differentiation of Human Mesenchymal Stem Cells

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Purpose: One of the design goals of ideal bone implant coating is osteoinductivity, the ability to induce osteogenic differentiation of human mesenchymal stem cells (hMSC). We evaluated the *in vitro* osteoinductive potential of high velocity oxy-fuel (HVOF) thermal sprayed TiO₂-10wt% HA (TiO₂-HA) nanocomposite coatings as alternatives to conventional FDA approved atmospheric plasma spray (APS) HA coatings. The TiO₂-HA nanocomposite coatings exhibit at least 2X higher bond strength values on Ti64 substrates than those of APS HA coatings, offering higher stability post-implantation. It is hypothesized that TiO₂-HA nanocomposites optimal nano and micro surface morphology and hybrid TiO₂/HA chemistry will enhance hMSC osteoblastic differentiation.

Methods: Human MSCs were isolated from multiple adult bone marrow samples as per our prior method [1] with Research Ethics Committee approval. The isolated hMSC were culture-expanded in basal (DMEM/10% FBS) and osteogenic medium (DMEM/10% FBS supplemented with 100nM dexamethasone, 10mM glycerolphosphate and 50 μM ascorbic acid-2-phosphate). Following proliferation and osteogenic differentiation (hMSC-ob), the hMSC and hMSC-ob were seeded on TiO₂-HA and reference HA and TiO₂ coatings. At predetermined culture time points, hMSC and hMSC-ob cultures were evaluated for (i) proliferation (Alamar Blue); (ii) cytoskeleton organization (F-actin/Vinculin); (iii) cell-coatings: interaction, growth and morphology (SEM); (iv) osteoblastic activity: alkaline phosphatase (ALP), osteocalcin (OC) expression; and (v) cellular mineralization: Alizarin Red (ARS) and fluorescent Calcein stains. ANOVA was used to determine statistical significance with a p < 0.05. Details concerning coatings preparation and processing were presented elsewhere [2].

Results: Both hMSCs and hMSCs-ob cultures revealed 1.5 times higher cellular metabolism (Alamar Bleu) when plated on TiO₂-HA nanocomposite coatings as compared with TiO₂ and HA reference coatings. Similarly after 14 days, the cytoskeletal organization of hMSCs and hMSC-ob cultures demonstrated more intense, dense and widespread F-actin deposition on the TiO₂-HA nanocomposites than on reference coatings (Fig. 1). Additionally, the cell growth kinetics, represented by red nuclei, were increased when plated on TiO₂-HA nanocomposites coatings and unaffected by culture conditions (basal unsupplemented vs. osteogenic medium), indicating substrate dependant behaviour. The level of expression of the osteoblastic markers ALP and OC was significantly higher in cells cultured on TiO₂-HA nanocomposites at day 7 and 21, the respective ALP and OC temporal peak times (Fig. 2).

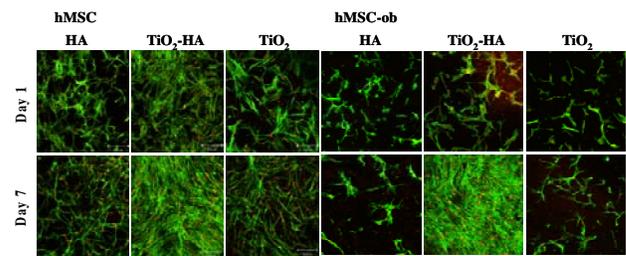


Figure 1. F-actin (green) organization and nuclei morphology (red) of hMSC and hMSC-ob at day 1 and 7 cultured on HA, TiO₂-HA nanocomposites and TiO₂ coatings. Original magnification was 200x.

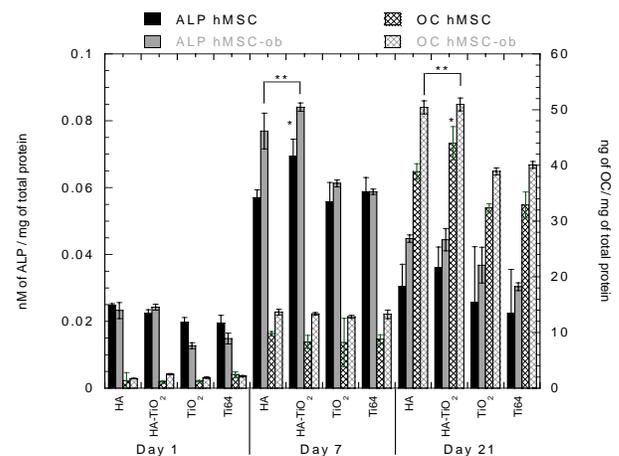


Figure 2: Normalized alkaline phosphatase activity (ALP), and osteocalcin expression (OC) in hMSC and hMSC-ob cells cultured on HA, TiO₂-HA nanocomposites and TiO₂ coatings and uncoated Ti64.

This trend of up-regulation of gene expression was also observed in the extracellular matrix mineral content as seen through Calcein and ARS histochemical staining. Indeed, the co-localization of ARS and Calcein stain in the absence of dexamethasone (hMSC cultures) was only observed when hMSCs were plated on TiO₂-HA. Control hMSC cultures on HA and TiO₂ were only found to mineralize when supplemented with dexamethasone. This suggests nodular cell aggregates are made up of cells committed to the osteoblastic lineage mainly driven by TiO₂-HA nanocomposite coating.

Conclusions: The present data demonstrated that the novel TiO₂-HA nanocomposite coatings support of the growth, proliferation, and differentiation of not only hMSCs, but also osteoblasts that derive from hMSCs. These findings collectively demonstrate the osteoinductivity potential of the nanostructured TiO₂-HA nanocomposite coatings and their potential as novel mechanically (bond strength) and biologically improved load bearing implants coatings.

References:

- [1] Mwale F. *et al.*, Tissue Eng. 2006;9:2639-47.
- [2] Lima R S *et al.*, J Thermal Spray Technol., 2007;16:40-63.