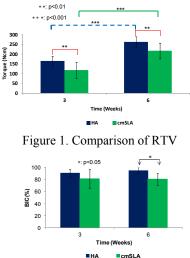
Effects of Hydroxyapatite-Coating vs. Hydrophilic Surface on Early Healing of Dental Implant in an Ovine Model Jin Whan Lee, Hai Bo Wen, Jeffrev Bassett

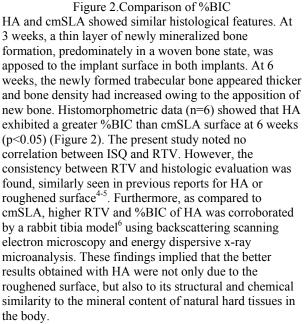
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Statement of Purpose: A pressurized, hydrothermal, post-plasma-sprayed hydroxyapatite (HA) coating has been used to enhance the biocompatibility and integration of dental implants. HA-coated implants healed more rapidly with a greater percentage of bone-to-implant contact (%BIC) than those of sand blasted with alumina and acid etched¹. Other documented benefits of HA coating in comparison to uncoated implant surfaces included significantly higher survival rates². Roughened surfaces can also achieve faster bone integration, as well as a higher %BIC and greater removal torque values (RTV) than machined surfaces³. One such surface is fabricated by sequentially sandblasting the surface with large grit, acid-etching the roughened surface, treatment with nitrogen gas, and final storage in a saline solution (cmSLA). However, evaluations seldom have been performed between cmSLA and other surfaces, such as HA. The objective of the study was to compare the bone tissue response and stability of implants with two different surfaces, cmSLA and HA coating during the early stages of wound healing in an ovine femoral condvle model.

Methods: Surgical Procedure: A total of 60 implants (4.1 mm x 10 mm) with MP1[®] HA-coated surface (Tapered Screw-Vent[®] implant with MP-1[®] HA surface, Zimmer) (n=30) and chemically modified, sand blasted and acidetched surface (SLActive[®] surface, Bone Level Implant, Straumann) (n=30) was randomly placed bilaterally in the femoral condules of ovine(6 implants per ovine). Implant stability Assessment: Implant stability quotients (ISQ) using resonance frequency analysis (Osstell® Mentor, Integration Diagnostics AB, Göteborg, Sweden) and mechanical torque device (Sturtevant Richmont, Carol Stream, IL) connected to data acquisition (MESURgauge, version 1.5, Mark-10, Copiague, NY) were measured at implant placement (as a baseline), 3 and 6 weeks of healing. Histologic and histomorphometric evaluation: After being dissected, specimen blocks were fixed in 10% formalin, dehydrated in ascending solutions of EtOH, infiltrated, and embedded in PMMA for undecalcified sectioning. Sections were cut (Makro Trennsystem; Exakt Apparatebau AG, Norderstedt, Germany), ground, and polished. The sections (≅50µm) were stained with toluidine blue and basic fuchsin, and examined using light microscopy (Olympus BH-2, Olympus Optical Company, Japan) to analyze %BIC. Statistical analysis: TheSatterthwaite method was used to compute the DF for F test and p values, followed by the Tukey post-hoc test. **Results:** There were no implant failures and all samples harvested at 3 and 6 weeks exhibited no adverse tissue responses. The difference in ISQ (n=15) values between 0 and 3 weeks was -0.5±6.6 for HA and 3.0±4.0 for cmSLA (p=0.3251). The difference in ISO (n=15) values between 0 and 6 weeks was 2.7 ± 7.1 for HA and 5.2 ± 5.9 for cmSLA (p=0.8409). No significant treatment effect (P>0.05) was found from the ISQ measurement. Greater

RTV (n=9) was found for implant with HA-coated surface as compared with cmSLA surface (p<0.01 at 3 or 6 weeks, respectively) (Figure 1). The torque values for HA were 39% and 21% higher than for cmSLA at 3 and 6 weeks, respectively.





Conclusions: A higher level of implant stability as demonstrated by RTV and bone apposition was achieved with an HA coated surface than a cmSLA surface during the early stage of healing for up to 6 weeks. **References:** 1.Eom TG. Oral Surg Oral Med Oral Pathol Oral Radiol 2012;114(4):411-8 2. Artzi Z. COIR 2006:17(1): 85-93 3. Abrahamsson I. Int J Oral Maxillofac Implants 2001:16(3):323-32 4. Svanborg LM. Int J Oral Maxillofac Surg 2011;40(3):308-15. 5. Klokkevold PR. Clin Oral Implants Res 2001;12(4):350-7. 6. Queiroz TP. Clin Oral Implants Res 2012, Epub.